#### Nitric Oxide 46 (2015) 25-31

Contents lists available at ScienceDirect

### Nitric Oxide

journal homepage: www.elsevier.com/locate/yniox

# H<sub>2</sub>S-releasing drugs: Anti-inflammatory, cytoprotective and chemopreventative potential

Burcu Gemici<sup>a</sup>, Wagdi Elsheikh<sup>b</sup>, Karla B. Feitosa<sup>c</sup>, Soraia K.P. Costa<sup>c</sup>, Marcelo N. Muscara<sup>c</sup>, John L. Wallace<sup>d,e,\*</sup>

<sup>a</sup> Near East University, Nicosia, Cyprus

<sup>b</sup> Department of Medicine, McMaster University, Hamilton, Ontario, Canada

<sup>c</sup> Department of Pharmacology, Institute of Biomedical Sciences, University of Sao Paulo, Av. Prof. Lineu Prestes, 1524, São Paulo 05508-000, SP, Brazil

<sup>d</sup> Department of Physiology and Pharmacology, University of Calgary, Calgary, Alberta, Canada

<sup>e</sup> Department of Pharmacology & Toxicology, University of Toronto, Toronto, Ontario, Canada

### ARTICLE INFO

Article history: Available online 18 November 2014

Keywords: Gastrointestinal Cytoprotection Inflammation Hydrogen sulfide Drug development

### ABSTRACT

Hydrogen sulfide exerts a number of cytoprotective and anti-inflammatory effects in many organ systems. In an effort to exploit these potent and beneficial effects, a number of hydrogen sulfide-releasing derivatives of existing drugs have been developed and extensively tested in pre-clinical models. In particular, efforts have been made by several groups to develop hydrogen sulfide-releasing derivatives of a number of nonsteroidal anti-inflammatory drugs. The main goal of this approach is to reduce the gastrointestinal ulceration and bleeding caused by this class of drugs, particularly when used chronically such as in the treatment of arthritis. However, these drugs may also have utility for prevention of various types of cancer. This paper provides an overview of some of the mechanisms underlying the anti-inflammatory and cytoprotective actions of hydrogen sulfide. It also gives some examples of hydrogen sulfidereleasing anti-inflammatory drugs, and their actions in terms of reducing inflammation and attenuating the development of cancer in experimental models.

© 2014 Elsevier Inc. All rights reserved.

### Contents

1.	Introduction	25
2.	Anti-inflammatory and pro-resolution effects of hydrogen sulfide	26
3.	Cytoprotective actions of hydrogen sulfide in GI tract	26
4.	GI-sparing anti-inflammatory drugs	27
5.	Chemopreventative actions of H <sub>2</sub> S-releasing NSAIDs	29
6.	Future directions	30
	References	30

E-mail address: altapharm@hotmail.com (J.L. Wallace).

### 1. Introduction

Over the past decade, several novel H<sub>2</sub>S-based drugs have been described [1,2]. Several of the most advanced such compounds are derivatives of currently used anti-inflammatory drugs. The H<sub>2</sub>S-releasing drugs show greatly reduced toxicity (particularly in the GI tract) and, in some cases, improved efficacy. Several nonsteroidal anti-inflammatory drugs (NSAIDs) have been found to substantially reduce the incidence of several types of human cancers, but their adverse effects preclude their use for these indications, which would require long-term exposure to the drugs. H<sub>2</sub>S itself exerts potent anti-inflammatory effects [3] and, in some test systems, anti-cancer effects [4,5]. In this article, we describe some of the features of H<sub>2</sub>S-releasing drugs that we have developed with respect



Review



Nitric Oxide

Abbreviations: ATP, adenosine triphosphate; COX, cyclooxygenase; CSE, cystathionine Υ-lyase; GI, gastrointestinal; ICAM, intercellular adhesion molecule; IFN, interferon; IL, interleukin; LFA, lymphocyte function-associated antigen; NFκB, nuclear factor κB; Nrf2, nuclear factor erythroid 2-related factor 2; NSAID, nonsteroidal antiinflammatory drug; PG, prostaglandin; TNF, tumour necrosis factor.

Declaration: Dr. Wallace is the founder and a director of Antibe Therapeutics Inc., which is developing  $H_2S$ -releasing anti-inflammatory drugs.

<sup>\*</sup> Corresponding author. 15 Prince Arthur Avenue, Toronto, Ontario, Canada M5R 1B2.

to their potential use as anti-inflammatory, analgesic and chemoprevention applications, as well as discussing the cytoprotective mechanisms of these drugs that so profoundly reduce their toxicity.

## 2. Anti-inflammatory and pro-resolution effects of hydrogen sulfide

Among the earliest events in an acute inflammatory reaction is the adherence of leukocytes to the vascular endothelium and their subsequent extravasation and migration to sites of injury. A key early step in this process is up-regulation of adhesion molecules on leukocytes and on the vascular endothelium. H<sub>2</sub>S plays a very important role in regulating these processes (Fig. 1). In fact, tonic production of H<sub>2</sub>S is an important contributor to the down-regulation of leukocyte adhesion in the healthy state. When an inhibitor of H<sub>2</sub>S synthesis is administered, there is a rapid increase in leukocyte adherence to the vascular endothelium [3]. This process is mediated via up-regulation of lymphocyte function-associated antigen (LFA) on circulating leukocytes and P-selectin and intercellular adhesion molecule (ICAM-1) on the endothelium [6] (Fig. 1). Rats with a diet-induced vitamin B deficiency, which have a diminished capacity to synthesize H<sub>2</sub>S, also exhibit significantly enhanced accumulation of leukocytes in the affected tissues [7]. Mice that are heterozygous for the cystathionine  $\beta$ -synthase gene also have diminished synthesis of H<sub>2</sub>S, and they exhibit increased levels of leukocyte adherence to the vascular endothelium, slower leukocyte rolling velocity and increased vascular permeability [8].

In contrast, a reduction of leukocyte adherence can be seen after administration of  $H_2S$  donors [3]. This effect appears to be



**Fig. 1.** Regulation of leukocyte–endothelial adherence by hydrogen sulfide (H<sub>2</sub>S). In health, H<sub>2</sub>S production by vascular smooth muscle or endothelial cells tonically down-regulates leukocyte adherence. Cystathionine Y-lyase (CSE) is likely the primary source of H<sub>2</sub>S synthesis in this setting. H<sub>2</sub>S acts via ATP-sensitive potassium channels (K<sub>ATP</sub>) to inhibit expression of adhesion molecules on the leukocytes (CD11/CD18) and endothelium (P-selectin, ICAM-1). When H<sub>2</sub>S synthesis is inhibited, such as by  $\beta$ -cyanolalanine (BCA; CSE inhibitor), up-regulation of leukocyte adherence to the endothelium, followed by extravasation of the leukocytes. Oedema formation also occurs during the extravasation process.

mediated by activation of adenosine triphosphate (ATP)-sensitive potassium channels on endothelial cells and leukocytes. H<sub>2</sub>S donors have also been shown to inhibit endothelial ICAM-1 expression triggered by high blood–glucose concentrations [9]. H<sub>2</sub>S donors have been shown to cause a marked suppression of inflammatory responses in several animal models, including colitis and endotoxic shock [10,11].

In addition to reducing leukocyte extravasation and migration to sites of injury, H<sub>2</sub>S can reduce the cytotoxic effects of neutrophils by inhibiting myeloperoxidase activity [12]. Moreover, H<sub>2</sub>S promotes apoptosis of neutrophils, a key step in resolution of inflammation [13]. H<sub>2</sub>S also increases phagocytosis of bacteria by macrophages, and promotes a shift of phenotype of macrophages to the "M2", pro-resolution state [14]. H<sub>2</sub>S donors can also reduce inflammation by reducing the expression of a number of proinflammatory cytokines (e.g., tumour necrosis factor (TNF)- $\alpha$ , interleukin (IL)-1β, IL-8, interferon (IFN)-Y but sparing of expression of the pro-inflammatory cytokine, IL-10 [10,11,15]. Inhibition of pro-inflammatory cytokine expression most likely occurs by suppression of nuclear factor KB (NFKB) activity by H<sub>2</sub>S [16]. We recently described a regulatory interaction between IL-10 and H<sub>2</sub>S [7]. IL-10-deficient mice exhibit impaired H<sub>2</sub>S synthesis in the colon (and these mice can spontaneously develop colitis) [7,17]. Administration of H<sub>2</sub>S donors increases IL-10 expression [7,15], while administration of IL-10 to the IL-10-deficient mice restored normal colonic H<sub>2</sub>S synthesis [7].

Some of the resolution-promoting effects of  $H_2S$  are mediated via induction of cyclooxygenase (COX)-2 expression, and the ensuing synthesis of anti-inflammatory eicosanoids such as prostaglandin (PG)E<sub>2</sub> [18,19]. In the GI tract, for example, inhibition of  $H_2S$  synthesis leads to a decrease in COX-2 expression, a decrease in PGE<sub>2</sub> synthesis, an increase in mucosal inflammation and an impairment of healing of damaged tissue [18–21]. Administration of  $H_2S$ donors has the opposite effects [20–22]. The augmentation of COX-2 expression by  $H_2S$  donors likely also contributes to their ability to accelerate healing of wounds and ulcers [20,23].

### 3. Cytoprotective actions of hydrogen sulfide in GI tract

The ability of H<sub>2</sub>S to reduce damage in the GI tract is striking, particularly since this cytoprotective effect extends to prevention of the severe injury and bleeding that can be induced in the small intestine by repeated administration of NSAIDs [6,11,22-26]. There are a number of mechanisms of action through which endogenous cytoprotective substances (e.g., PGs, nitric oxide) produce their beneficial effects [27] and considerable data have been generated in recent years with respect to the actions of H<sub>2</sub>S in this regard (Fig. 2). For example, H<sub>2</sub>S will stimulate secretion of bicarbonate in the stomach and duodenum, thereby reducing the potentially damaging effects of gastric acid [24,28]. H<sub>2</sub>S may also directly inhibit gastric acid secretion [29]. A reduction in gastric blood flow is a common feature in the early stages of gastric injury, and it can be prevented by H<sub>2</sub>S donors [6]. Likewise, gastric damage induced by NSAIDs is largely mediated, particularly at the earliest stages, by leukocyte adhesion to the vascular endothelium in the gastric microcirculation [30].

As mentioned earlier, the ulceration and bleeding caused by NSAIDs in the small intestine is much more complex, in terms of pathogenesis, and develops over a longer period of time than the damage in the stomach [31,32]. Unlike damage in the stomach and duodenum, gastric acid does not play a significant role in the damage induced by NSAIDs in the more distal small intestine. Indeed, the use of proton pump inhibitors and histamine H<sub>2</sub>-receptor antagonists to reduce NSAID-induced damage in the upper GI tract results in exacerbation of ulceration and bleeding in the small intestine [33,34]. This is a consequence of significant changes in the



**Fig. 2.** Mechanisms for cytoprotective effects of hydrogen sulfide (H<sub>2</sub>S) in the gastrointestinal tract. In the stomach, the ability of H<sub>2</sub>S to maintain blood flow in the face of injurious agents (such as nonsteroidal anti-inflammatory drugs), stimulation of bicarbonate secretion and inhibition of leukocyte adherence to the vascular endothelium all contribute to increased resistance to ulceration. H<sub>2</sub>S can also accelerate healing of gastric ulcers after they have formed. Protection of the small intestine by H<sub>2</sub>S likely involves multiple mechanisms, including prevention of uncoupling of oxidative phosphorylation (thereby preventing mitochondrial injury) and reducing the cytotoxic effects of bile. The latter may occur in part through the ability of H<sub>2</sub>S to alter the composition of the intestinal microbiota. Increased mucus production by intestinal goblet cells and increased expression of cyclo-oxygenase (COX)-2, which produces prostaglandin E<sub>2</sub> and promotes repair, further contribute to the cytoprotective actions of H<sub>2</sub>S.

intestinal microbiota when gastric acid secretion is suppressed, with a marked diminution of levels of *Bifidobacteria* [33].

The mechanisms through which H<sub>2</sub>S can protect against NSAIDenteropathy appear to include changes in at least three of the key factors that are central to the pathogenesis of this injury: the composition/secretion of bile, the microbiota and the enterohepatic recirculation of NSAIDs [22]. Thus, H<sub>2</sub>S reduces the cytotoxicity of bile (which is elevated by NSAIDs), promotes a normalization of the microbiota (which is altered by NSAIDs) and reduces the enterohepatic circulation of NSAIDs and/or NSAID-glucuronides [22,24,35].

### 4. GI-sparing anti-inflammatory drugs

NSAIDs are among the most commonly used drugs, because they effectively reduce pain and inflammation. The use of this class of drugs on a chronic basis is for joint inflammation, most notably osteoarthritis. While very effective, and non-addictive, NSAIDs do carry a significant risk for adverse effects, with the most clinically significant being GI ulceration and bleeding. Unfortunately, selective COX-2 inhibitors provided only a partial solution to this problem, and their use is associated with significant cardiovascular toxicity [36]. H<sub>2</sub>S appears to provide a solution to these problems. A range of NSAID derivatives that release H<sub>2</sub>S have been synthesized and evaluated, with consistent results in terms of retaining anti-inflammatory activity but reducing GI toxicity. For example, we have synthesized derivatives of aspirin, naproxen, ketoprofen,

indomethacin, ibuprofen, flurbiprofen and diclofenac, each with a number of different H<sub>2</sub>S-releasing moieties. Data are presented in this paper on aspirin, naproxen and ketoprofen derivatives, as representative examples. The major focus of our research has been on a compound called ATB-346, which consists of naproxen covalently linked to 4-hydroxythiobenzamide (TBZ) [26,34]. We have been studying this drug as a potential treatment for arthritis, a chronic use. Figure 3 shows the effects of ATB-346 and naproxen in a rat model in which arthritis was induced by injection of Freund's adjuvant [37]. ATB-346 produced a dose-dependent reduction of paw oedema in this model, not significantly different from the effects of equimolar doses of naproxen. GI damage was frequently observed in the naproxen-treated rats, but not in the rats treated with ATB-346. Indeed, at the highest dose tested, all of the 12 rats treated with naproxen had died before completion of the study because of perforation of the small intestine (Fig. 3).

We chose naproxen as the NSAID to use as the base drug because it is the only marketed NSAID that has not been associated with significant and serious cardiovascular adverse events [36]. Indeed, concerns about these cardiovascular events have led to several large physician's groups (e.g., American Heart Association, American College of Gastroenterology) to recommend use of naproxen, or of a selective COX-2 inhibitor plus low-dose aspirin (the latter for cardioprotection). However, co-administration of low-dose aspirin with NSAIDs (including selective COX-2 inhibitors) greatly increases the risk of GI ulceration and bleeding [38]. A further recommendation in many patients is that they take drugs to suppress gastric acid secretion (e.g., proton pump inhibitors, histamine H<sub>2</sub> receptor antagonists) to protect their stomach from damage, but these drugs significantly worsen ulceration and bleeding in the small intestine [33], where it is more difficult to detect and for which there are no proven-effective preventative or curative treatments [31,32,39,40].

The metabolism of ATB-346 and the mechanism for  $H_2S$  release from this drug are not completely understood. Release of  $H_2S$  from ATB-346 has been demonstrated *in vitro* [41]. The release of  $H_2S$  from this compound occurs at a very low level when the drug is dissolved in a buffer, but is greatly increased in the presence of tissue or in the presence of reducing agents such as dithiothreitol, L-cysteine



**Fig. 3.** Dose-dependent reduction of adjuvant-induced arthritis in rats by naproxen and ATB-346. Twice-daily treatment with either drug produced a dose-dependent reduction of paw swelling. However, at the highest dose tested, all rats treated with naproxen were moribund (requiring euthanasia) or had died by day 7 (arrow) as a result of perforated ulcers in the intestine. Gastrointestinal damage was not observed in the rats treated with ATB-346. Results are shown as the mean ± SEM, with 12 rats per group.



**Fig. 4.** Proposed mechanism of release of hydrogen sulfide ( $H_2S$ ) and naproxen from ATB-346 after oral administration. Biological effects consistent with  $H_2S$  release are evident within minutes of administration of ATB-346 (e.g., gastric vasodilation). While suppression of gastric prostaglandin synthesis and whole blood thromboxane synthesis can be observed within 15 minutes of administration of ATB-346, a significant rise in blood levels of naproxen only occurs over several hours.

or glutathione. One likely scenario of metabolism of ATB-346 is shown in Fig. 4. Hydrolysis of the thiol moiety results in H<sub>2</sub>S release, leaving naproxen-4-hydroxybenzamide. This compound suppresses COX activity and induces gastric damage when given to rats

(Figs 4, 5). As shown in Fig. 5, the rise in plasma naproxen levels in rats after oral administration of ATB-346 is guite slow as compared to that observed after oral administration of naproxen itself. However, there is significant suppression of gastric prostaglandin synthesis within 15 min of administration of ATB-346, when plasma levels of naproxen are very low. This supports the notion that an intermediate with COX-inhibitory activity is rapidly formed after oral administration of ATB-346. The lack of gastric damage observed following oral administration of ATB-346 (Fig. 4) is most likely due to the rapid release of H<sub>2</sub>S. We have confirmed that H<sub>2</sub>S donors can replicate this protective effect, and that protection of the small intestine from injury is also achieved [18,22]. The latter is significant because the enteropathy caused by NSAIDs is much more slow to develop than the gastric damage, so one might predict that if H<sub>2</sub>S was rapidly released from ATB-346, any beneficial effects might not be long-lasting enough to protect the small intestine. Our pharmacokinetic studies demonstrate that the total exposure of ATB-346derived naproxen from oral dosing with this drug, as compared to dosing with naproxen itself at an equimolar dose, is very similar (~14% lower bioavailability with ATB-346). We have also observed that there is significantly reduced enterohepatic recirculation of naproxen following administration of ATB-346. Biliary levels of naproxen and naproxen-glucuronides were markedly reduced following ATB-346 administration to rats, as compared to rats treated with naproxen itself [34]. It is possible that this effect is also related to alterations in the microbiota, given that bacterial enzymes account for deglucuronidation of NSAID-glucuronides in the intestine [42].

The lack of GI injury in rats treated with ATB-346 is impressive, but there are NSAIDs marketed for treatment of arthritis that are much more toxic in the GI tract than naproxen, and these drugs have particular utility for treatment of acute, severe pain (post-surgery, gout, etc.). Ketoprofen is one such NSAID. We examined the GI-sparing effects of ATB-352, an H<sub>2</sub>S-releasing derivative of ketoprofen, in rats (6 per group). Oral administration of ketoprofen



**Fig. 5.** Pharmacokinetics and pharmacodynamics of ATB-346. Inhibition of gastric prostaglandin synthesis (and whole blood thromboxane synthesis; not shown) is apparent within 15 min of oral administration of ATB-346 to rats (panel A), but blood levels of naproxen are very low (panel B), suggesting that ATB-346 itself or a rapidly generated metabolite can inhibit cyclooxygenase activity. Release of hydrogen sulfide (H<sub>2</sub>S) from ATB-346 via hydrolysis results in the formation of naproxen-4-hydroxybenzamide, which can inhibit cyclooxygenase. When administered to rats, naproxen-4-hydroxybenzamide produced significant gastric hemorrhagic damage (panel C), while ATB-346 administration did not induce gastric damage (panel D). Results in panels A and B are shown as the mean ± SEM, with 6 rats per group.

at a dose of 30 mg/kg to fasted rats resulted, within a few hours, in the formation of extensive haemorrhagic erosions in the stomach. The mean gastric damage score was  $41 \pm 3$ . In contrast, oral administration of an equimolar dose of ATB-352 did not produce any gastric damage (confirmed by histology). As has been observed with other H<sub>2</sub>S-releasing NSAIDs, ATB-352 inhibited gastric PGE<sub>2</sub> synthesis as effectively as ketoprofen.

To determine if similar safety of ATB-352 would be observed in the small intestine, groups of 6 rats each were treated orally with ketoprofen (20 mg/kg) or an equimolar dose of ATB-352 twice daily for 5 days (the rats were not fasted). Treatment with ketoprofen elicited extensive ulceration and bleeding in the intestine, with a mean damage score of over 700 (Fig. 6). This level of damage is approximately 7 times that observed with naproxen administered at 20 mg/ kg. In sharp contrast, administration of an equimolar dose of ATB-352 did not provoke significant intestinal damage (mean score of 2; same as in vehicle-treated rats), despite inhibiting small intestinal PGE<sub>2</sub> synthesis and whole blood thromboxane synthesis as effectively as ketoprofen (by >90%; Fig. 6).

Chronic use of aspirin has been shown to reduce the incidence of a range of serious cardiovascular events, as well as reducing the incidence of several types of cancer [43–45]. The primary limitation to such use is the significant risk of aspirin-induced bleeding, particularly in the GI tract. A compound that showed the desirable effects of aspirin without the GI adverse effects would have tremendous potential for chronic use as a chemopreventative agent. With this in mind, we evaluated the properties of a novel H<sub>2</sub>S-releasing derivative of aspirin, ATB-340. The beneficial effects of aspirin with respect to cardiovascular disease prevention are attributable to the ability of this drug to irreversibly inhibit COX-1, particularly in platelets. Thus, we first determined if ATB-340 exhibited comparable inhibitory activity on COX-1 activity in an in vitro model. Human blood samples were added to tubes containing vehicle, aspirin or ATB-340 at concentrations of 1-30 µM. The blood was allowed to clot at room temperature for 45 min, after which the tubes were centrifuged and the serum was collected for measurement of thromboxane B<sub>2</sub> levels by enzyme-linked immunosorbent assay [46]. Both aspirin and ATB-340 inhibited thromboxane synthesis in a concentration-dependent manner (>95% inhibition at 30 µM), with no significant differences observed between the two drugs. We then proceeded to evaluate ATB-340 *in vivo*. Groups of rats (n = 6 each) were fasted overnight and then given vehicle or one of the two test drugs orally, at doses ranging from 3 to 100 mg/kg. Three hours later, the rats were euthanised and the stomach was examined by an observer blind as to the treatments the rats had received. Haemorrhagic gastric damage was



**Fig. 7.** Gastric damaging effects of 1 week of daily administration of aspirin or ATB-340 to rats. Aspirin at 10 mg/kg caused significant hemorrhagic damage in the stomach (\*\*p < 0.01 vs. vehicle-treated), while ATB-340 did not produce damage at either dose. Each group consisted of 6 rats (mean ± SEM; ANOVA and Dunnett's multiple comparison test).

scored (the area of each lesion was measured in mm<sup>2</sup> and these were summed to give the total area of all lesions in the stomach). Aspirin caused haemorrhagic damage that increased in severity with dose, while damage was not seen in the rats treated with ATB-340 at equimolar doses. Thus, the level of damage in rats treated with ATB-340 did not differ significantly from that in vehicle-treated rats. We then examined the effects of repeated administration of aspirin vs. ATB-340 in rats. As shown in Fig. 7, administration of aspirin each day for a week at a dose of 10 mg/kg resulted in significant damage to the stomach. In contrast, with daily administration of ATB-340, no significant gastric damage was observed (Fig. 7).

### 5. Chemopreventative actions of H<sub>2</sub>S-releasing NSAIDs

There is considerable evidence that regular use of NSAIDs, including aspirin, can significantly reduce the incidence of a number of types of cancer [47,48], and they appear to be particularly effective at reducing the incidence of colon cancer [43,44]. However, their propensity to cause ulceration and bleeding is a major barrier to their use for chemoprevention of cancers (or for other chemoprevention uses). We and others have explored the possibility that GI-sparing, H<sub>2</sub>S-releasing NSAIDs would produce the beneficial chemopreventative effects without the GI toxicity [49,50]. For example, we evaluated the effects of a H<sub>2</sub>S-releasing ketoprofen derivative, ATB-352, in a model of pre-cancerous lesions in mice. Five-week-old mice were given injections of azoxymethane (10 mg/kg, i.p.) at weekly intervals (total of 4 injections). The mice were



**Fig. 6.** Twice-daily administration of ketoprofen (20 mg/kg) to rats resulted in extensive small intestinal ulceration, while rats treated with an equimolar dose of ATB-352 did not develop intestinal damage (left panel). The lack of intestinal damage with ATB-352 was observed despite comparable suppression of intestinal prostaglandin synthesis (middle panel) or whole blood thromboxane synthesis (right panel). The results are shown as the mean  $\pm$  SEM, with 6 rats per group (\*\*\*p < 0.001 vs. the other groups; ANOVA and Dunnett's multiple comparison test).

euthanised 1 week after the final injection of azoxymethane. A laparotomy was performed and the entire colon was excised. After gentle flushing with 0.9% saline, the colon was tied at both ends with silk sutures and insufflated with 10% phosphate-buffered formalin. The colons were submerged in formalin for 24 h, then stained with 0.2% methylene blue. Using a dissection microscope at 40× magnification, the number of aberrant crypt foci (ACF) in the entire colon was blindly determined. ACFs are pre-cancerous lesions which were distinguishable from the surrounding normal crypts by their increased size, significantly increased distance from the lamina to basal surface and easily discernible pericryptal zone [51]. Quantification of ACF was performed blindly, according to previously published criteria [52].

Groups of at least 6 rats each were treated once-daily for the first 2 weeks of azoxymethane administration with vehicle, ketoprofen at 10 mg/kg, or equimolar doses of ATB-352 or of the H<sub>2</sub>S-releasing moiety of ATB-352 (TBZ). As shown in Fig. 8, vehicle-treated rats developed an average of ~35 aberrant crypt foci. Treatment with ketoprofen or ATB-352 significantly reduced this number by 40–50%, with no significant difference between these two drugs. The two drugs also inhibited colonic PGE<sub>2</sub> synthesis to a similar extent (>90% inhibition). Treatment with TBZ had no significant effect on numbers of aberrant crypt foci, and did not significantly affect colonic PGE<sub>2</sub> synthesis.

We further evaluated the chemopreventative effects of ATB-352 in mice with a genetic defect that predisposes them to intestinal cancer [53]. Male Apc<sup>Min/+</sup> mice were obtained from Jackson Laboratories (Bar Harbor, MA, USA). The mice were treated with vehicle, ketoprofen at 10 mg/kg, or equimolar doses of ATB-352 or TBZ once-daily for 2 weeks (beginning at 6 weeks of age). When the mice reached week 14 of age, the colon and small intestine were examined for polyps. The area (in mm<sup>2</sup>) of polyps was measured under a dissecting microscope as described previously, by an observer unaware of the treatments the mice had received [54]. The total area of all polyps was calculated as the "polyp score".

Figure 9 shows the effects of the drugs on the polyp score. In vehicle-treated mice, the average polyp score was ~100. Treatment with ketoprofen significantly reduced the polyp score by ~50% (p < 0.05). Treatment with ATB-352 was significantly more effective than treatment with ketoprofen (~60% reduction). However, TBZ had no significant effect.



**Fig. 8.** Daily treatment of mice with ketoprofen (10 mg/kg) or an equimolar dose of a hydrogen sulfide-releasing derivative of ketoprofen (ATB-352) significantly reduced the numbers of aberrant crypt foci in mice that had received the carcinogen azoxymethane (p < 0.001 vs. vehicle-treated; n = 8 per group). Treatment with the hydrogen sulfide-releasing moiety of ATB-352 (4-hydroxythiobenzamide; TBZ) had no significant effect.



**Fig. 9.** The number of intestinal polyps that spontaneously developed in Apc<sup>Min/+</sup> mice was significantly reduced by daily treatment for 2 weeks with ketoprofen (10 mg/ kg) or with an equimolar dose of ATB-352, a hydrogen sulfide-releasing derivative of ketoprofen (\*p < 0.05, \*\*p < 0.01 vs. the vehicle-treated group). The effect of ATB-352 was significantly greater than the effect of ketoprofen (\* $\psi < 0.05$ ). Treatment with the hydrogen sulfide-releasing moiety of ATB-352 (TBZ; 4-hydroxythiobenzamide) had no significant effect on the numbers of polyps that developed. The rats were treated during the 6th and 7th weeks of age, and the polyps were blindly measured at 14 weeks of age.

These results in animal models suggest that H<sub>2</sub>S-releasing NSAIDs may be effective and safe in chemoprevention of colon cancer, and other types of cancers. The mechanism of action of these drugs is not clear, but suggested mechanisms for the anti-cancer actions of H<sub>2</sub>S include inactivation of Nrf2 (nuclear factor erythroid 2-related factor 2) via sulhydration of Keap1 [55], induction of glycolysis within cancer cells [5] and induction of apoptosis of cancer cells [56]. Inhibition of COX-2 activity could certainly be a contributing effect to the reduction of polyps and tumours, but the enhanced activity of ATB-352 as compared to ketoprofen is unlikely due to different effects on COX-2 activity. Our data demonstrated that these two drugs inhibit colonic PGE<sub>2</sub> synthesis to the same extent.

### 6. Future directions

Proof-of-concept animal studies of H<sub>2</sub>S-releasing anti-inflammatory drugs are very encouraging, particularly as such consistent effects have been observed across a range of anti-inflammatory drugs and a range of H<sub>2</sub>S-releasing moieties. These compounds exhibit profoundly increased safety profiles, particularly with respect to GI damage. They also often exhibit enhanced anti-inflammatory activity. NSAIDs are used primarily for treating acute and chronic pain, but as discussed in this paper, they are potentially very useful for a number of other indications, such as chemoprevention of cancer. The toxicity of NSAIDs is the major barrier to their widespread use in chemoprevention. Thus, the marked reduction of toxicity through linking of NSAIDs to H<sub>2</sub>S-releasing moieties opens up the possibility for much wider and longer-term use of these drugs.

Clinical studies of several  $H_2S$ -releasing drugs are now underway. With the advancement of such drugs into human studies, the need for improved methods for detection and measurement of  $H_2S$  *in vivo* will become even more urgent. Moreover, there will undoubtedly be new avenues for therapeutic exploitation of the powerful anti-inflammatory, chemopreventative cytoprotective effects of  $H_2S$  [5,19].

### References

- M.V. Chan, J.L. Wallace, Hydrogen sulfide-based therapeutics and gastrointestinal diseases: translating physiology to treatments, Am. J. Physiol. Gastrointest. Liver Physiol. 305 (2013) G467–G473.
- [2] G. Caliendo, G. Cirino, V. Santagada, J.L. Wallace, Synthesis and biological effects of hydrogen sulfide (H<sub>2</sub>S): development of H<sub>2</sub>S-releasing drugs as pharmaceuticals, J. Med. Chem. 53 (2010) 6275–6286.

- [3] R.C.O. Zanardo, V. Brancaleone, E. Distrutti, S. Fiorucci, G. Cirino, J.L. Wallace, Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation, FASEB J. 20 (2006) 2118–2120.
- [4] H.B. Ma, S. Huang, X.R. Yin, Y. Zhang, Z.L. Di, Apoptotic pathway induced by diallyl trisulfide in pancreatic cancer cells, World J. Gastroenterol. 20 (2014) 193–203.
- [5] Z.W. Lee, X.Y. Teo, E.Y. Tay, C.H. Tan, T. Hagen, P.K. Moore, et al., Utilizing hydrogen sulfide as a novel anti-cancer agent by targeting cancer glycolysis and pH imbalance, Br. J. Pharmacol. 171 (2014) 4322–4336.
- [6] S. Fiorucci, E. Antonelli, E. Distrutti, G. Rizzo, A. Mencarelli, S. Orlandi, et al., Inhibition of hydrogen sulfide generation contributes to gastric injury caused by anti-inflammatory nonsteroidal drugs, Gastroenterology 129 (2005) 1210– 1224.
- [7] K.L. Flannigan, T.A. Agbor, R.W. Blackler, J.J. Kim, W.I. Khan, E.F. Verdu, et al., Impaired hydrogen sulfide synthesis and IL-10 signaling underlie hyperhomocysteinemia-associated exacerbation of colitis, Proc. Natl Acad. Sci. U.S.A. 111 (2014) 13559–13564.
- [8] A.F. Kamath, A.K. Chauhan, J. Kisucka, V.S. Dole, J. Loscalzo, D.E. Handy, et al., Elevated levels of homocysteine compromise blood-brain barrier integrity in mice, Blood 107 (2006) 591–593, s.
- [9] Q. Guan, X. Wang, L. Gao, J. Chen, Y. Liu, C. Yu, et al., Hydrogen sulfide suppresses high glucose-induced expression of intercellular adhesion molecule-1 in endothelial cells, J. Cardiovasc. Pharmacol. 62 (2013) 278–284.
- [10] S. Fiorucci, S. Orlandi, A. Mencarelli, G. Caliendo, V. Santagada, E. Distrutti, et al., Enhanced activity of a hydrogen sulphide-releasing derivative of mesalamine (ATB-429) in a mouse model of colitis, Br. J. Pharmacol. 150 (2007) 996–1002.
- [11] L. Li, G. Rossoni, A. Sparatore, L.C. Lee, P. Del Soldato, P.K. Moore, Antiinflammatory and gastrointestinal effects of a novel diclofenac derivative, Free Radic. Biol. Med. 42 (2007) 706–719.
- [12] Z. Palinkas, P.G. Furtmuller, A. Nagy, C. Jakopitsch, K.F. Pirker, M. Magierowski, et al., Interactions of hydrogen sulfide with myeloperoxidase, Br. J. Pharmacol. (2014), doi:10.1111/bph.12769.
- [13] C.N. Serhan, S.D. Brain, C.D. Buckley, D.W. Gilroy, C. Haslett, L.A.J. O'Neill, et al., Resolution of inflammation: state of the art, definitions and terms, FASEB J. 21 (2007) 325–332.
- [14] N. Dufton, J. Natividad, E.F. Verdu, J.L. Wallace, Hydrogen sulfide and resolution of acute inflammation: a comparative study utilizing a novel fluorescent probe, Scientific Rep. 2 (2012) 499.
- [15] O. Zayachkivska, O. Havryluk, N. Hrycevych, N. Bula, O. Grushka, J.L. Wallace, Cytoprotective effects of hydrogen sulfide in novel rat models of non-erosive esophagitis, PLoS ONE 9 (2014) e110688.
- [16] N. Sen, B.D. Paul, M.M. Gadalla, A.K. Mustafa, T. Sen, R. Xu, et al., Hydrogen sulfide-linked sulfhydration of NFkB mediates its antiapoptotic actions, Mol. Cell 45 (2012) 13–24.
- [17] N.J. Davidson, M.W. Leach, M.M. Fort, L. Thompson-Snipes, R. Kühn, W. Müller, et al., T helper cell 1-type CD4+ T cells, but not B cells, mediate colitis in interleukin 10-deficient mice, J. Exp. Med. 184 (1996) 241–251.
- [18] J.L. Wallace, Physiological and pathophysiological roles of hydrogen sulfide in the gastrointestinal tract, Antiox. Redox Signal. 12 (2010) 1125–1133.
- [19] J.L. Wallace, R.W. Blackler, M.V. Chan, G.J. da Silva, K.L. Flannigan, I. Gamaniek, et al., Anti-inflammatory and cytoprotective actions of hydrogen sulfide: translation to therapeutics, Antioxid. Redox Signal. (2014), PMID: 24635322.
- [20] J.L. Wallace, M. Dicay, W. McKnight, G.R. Martin, Hydrogen sulfide enhances ulcer healing in rats, FASEB J. 21 (2007) 4070–4076.
- [21] J.L. Wallace, L. Vong, W. McKnight, M. Dicay, G.R. Martin, Endogenous and exogenous hydrogen sulfide promotes resolution of colitis in rats, Gastroenterology 137 (2009) 569–578.
- [22] R.W. Blackler, J.P. Motta, A. Manko, M. Workentine, P. Bercik, M.G. Surette, et al., Hydrogen sulfide protects against NSAID-enteropathy through modulation of bile and the microbiota, Br. J. Pharmacol. (2014), doi:10.1111/bph.12961.
- [23] A. Papapetropoulos, Z. Pyriochou, G. Altaany, A. Yang, Z. Marazioti, M.G. Zhou, et al., Hydrogen sulfide is an endogenous stimulator of angiogenesis, Proc. Natl Acad. Sci. U.S.A. 106 (2009) 21972–21977.
- [24] R.W. Blackler, B. Gemici, A. Manko, J.L. Wallace, NSAID-gastroenteropathy: new aspects of pathogenesis and prevention, Curr. Opin. Pharmacol. 19 (2014) 11–16.
- [25] J.L. Wallace, G. Caliendo, V. Santagada, G. Cirino, S. Fiorucci, Gastrointestinal safety and anti-inflammatory effects of a hydrogen sulphide-releasing diclofenac derivative in the rat, Gastroenterology 132 (2007) 261–271.
- [26] J.L. Wallace, G. Caliendo, V. Santagada, G. Cirino, Markedly reduced toxicity of a hydrogen sulfide-releasing derivative of naproxen (ATB-346), Br. J. Pharmacol. 159 (2010) 1236–1246.
- [27] J.L. Wallace, Prostaglandins, NSAIDs, and gastric mucosal protection: why doesn't the stomach digest itself?, Physiol. Rev. 88 (2008) 1547–1565.
- [28] K. Takeuchi, E. Aihara, M. Kimura, K. Dogishi, T. Hara, S. Hayashi, Gas mediators involved in modulating duodenal HCO<sub>3</sub><sup>-</sup> secretion, Curr. Med. Chem. 19 (2012) 43–54.
- [29] S.A. Mard, H. Askari, N. Neisi, A. Veisi, Antisecretory effect of hydrogen sulfide on gastric acid secretion and the involvement of nitric oxide, BioMed Res. Int. 2014 (2014) 480921, doi:10.1155/2014/480921.

- [30] J.L. Wallace, C.M. Keenan, D.N. Granger, Gastric ulceration induced by nonsteroidal anti-inflammatory drugs is a neutrophil-dependent process, Am. J. Physiol. 259 (1990) G462–G467.
- [31] J.L. Wallace, NSAID gastropathy and enteropathy: distinct pathogenesis likely necessitates distinct prevention strategies, Br. J. Pharmacol. 165 (2012) 67–74.
- [32] J.L. Wallace, Mechanisms, prevention and clinical implications of nonsteroidal anti-inflammatory drug-enteropathy, World J. Gastroenterol. 19 (2013) 1861–1876.
- [33] J.L. Wallace, S. Syer, E. Denou, G. de Palma, L. Vong, W. McKnight, et al., Proton pump inhibitors exacerbate NSAID-induced small intestinal injury by inducing dysbiosis, Gastroenterology 141 (2011) 1314–1322.
- [34] R. Blackler, S. Syer, M. Bolla, E. Ongini, J.L. Wallace, Gastrointestinal-sparing effects of novel NSAIDs in rats with compromised mucosal defence, PLoS ONE 7 (2012) e35196.
- [35] J.P. Motta, K.L. Flannigan, T.A. Agbor, J.K. Beatty, R.W. Blackler, M.L. Workentine, et al., Hydrogen sulfide protects from colitis and restores intestinal microbiota biofilm and mucus production, Inflamm. Bowel Dis, (2014) in press.
- [36] P.M. Kearney, C. Baigent, J. Godwin, H. Halls, J.R. Emberson, C. Patrono, Do selective cyclo-oxygenase-2 inhibitors and traditional non-steroidal antiinflammatory drugs increase the risk of atherothrombosis? Meta-analysis of randomised trials, BMJ 332 (2006) 1302–1308.
- [37] D.M. McCafferty, D.N. Granger, J.L. Wallace, Indomethacin-induced gastric injury and leukocyte adherence in arthritic versus healthy rats, Gastroenterology 109 (1995) 1173–1180.
- [38] C. Sostres, A. Lanas, Gastrointestinal effects of aspirin, Nat. Rev. Gastroenterol. Hepatol. 8 (2011) 385–394.
- [39] A. Lanas, L.A. García-Rodríguez, M. Polo-Tomás, M. Ponce, I. Alonso-Abreu, M.A. Perez-Aisa, et al., Time trends and impact of upper and lower gastrointestinal bleeding and perforation in clinical practice, Am. J. Gastroenterol. 104 (2009) 1633–1641.
- [40] J.L. Wallace, Polypharmacy of osteoarthritis: the perfect intestinal storm, Dig. Dis. Sci. 58 (2013) 3088–3093.
- [41] J.L. Wallace, G. Cirino, V. Santagada, G. Caliendo, Hydrogen sulfide derivatives of nonsteroidal anti-inflammatory drugs, United States Patent Application No. WO/2008/009127 (2008).
- [42] A. LoGuidice, B.D. Wallace, L. Bendel, M.R. Redinbo, U.A. Boelsterli, Pharmacologic targeting of bacterial β-glucuronidase alleviates nonsteroidal anti-inflammatory drug-induced enteropathy in mice, J. Pharmacol. Exp. Ther. 341 (2012) 447–454.
- [43] C. Patrono, B. Rocca, Aspirin: promise and resistance in the new millennium, Arteriosclerosis Thromb. Vasc. Biol. 28 (2008) S25–S32.
- [44] R.S. Sandler, S. Halabi, J.A. Baron, S. Budinger, E. Paskett, R. Keresztes, et al., A randomized trial of aspirin to prevent colorectal adenomas in patients with previous colorectal cancer, N. Engl. J. Med. 348 (2003) 883–890.
- [45] R.F. Logan, J. Little, P.G. Hawtin, J.D. Hardcastle, Effect of aspirin and non-steroidal anti-inflammatory drugs on colorectal adenomas: case-control study of subjects participating in the Nottingham faecal occult blood screening programme, BMJ 307 (1993) 285–289.
- [46] J.R. Goodman, D. Grossman, Aspirin and other NSAIDs as chemoprevention agents in melanoma, Cancer Prev. Res. 7 (2014) 557–564.
- [47] E. Yiannakopoulou, Targeting epigenetic mechanisms and microRNAs by aspirin and other nonsteroidal anti-inflammatory agents – implications for cancer treatment and chemoprevention, Cell. Oncol. 37 (2014) 167–178.
- [48] J.L. Wallace, A. Bak, G.W. McKnight, S. Asfaha, K.A. Sharkey, W.K. MacNaughton, Cyclooxygenase 1 contributes to inflammatory responses in rats and mice: implications for gastrointestinal toxicity, Gastroenterology 115 (1998) 101–109.
- [49] K. Kashfi, Anti-cancer activity of new designer hydrogen sulfide-donating hybrids, Antioxid. Redox Signal. 20 (2014) 831–846.
- [50] W. Elsheikh, R.W. Blackler, K.L. Flannigan, J.L. Wallace, Enhanced chemopreventative effects of a hydrogen sulfide-releasing anti-inflammatory drug (ATB-346) in experimental colorectal cancer, Nitric Oxide 41 (2014) 131–137.
- [51] J.M. Ward, R.S. Yamamoto, C.A. Brown, Pathology of intestinal neoplasms and other lesions in rats exposed to azoxymethane, J. Nat. Cancer Inst. 51 (1973) 1029–1039.
- [52] E.A. McLellan, R.P. Bird, Specificity study to evaluate induction of aberrant crypts in murine colons, Cancer Res. 48 (1988) 6183–6186.
- [53] A.R. Moser, C. Luongo, K.A. Gould, M.K. McNeley, A.R. Shoemaker, W.F. Dove, ApcMin: a mouse model for intestinal and mammary tumorigenesis, Eur. J. Cancer 31A (1995) 1061–1064.
- [54] S.J. Baek, R. Okazaki, S.H. Lee, J. Martinez, J.S. Kim, K. Yamaguchi, et al., Nonsteroidal anti-inflammatory drug-activated gene-1 over expression in transgenic mice suppresses intestinal neoplasia, Gastroenterology 131 (2006) 1553–1560.
- [55] G. Yang, K. Zhao, Y. Ju, S. Mani, Q. Cao, S. Puukila, et al., Hydrogen sulfide protects against cellular senescence via S-sulfhydration of Keap1 and activation of Nrf2, Antiox. Redox Signal. 18 (2013) 1906–1919.
- [56] H.B. Ma, S. Huang, X.R. Yin, Y. Zhang, Z.L. Di, Apoptotic pathway induced by diallyl trisulfide in pancreatic cells, World J. Gastroenterol. 20 (2014) 193–203.